

## CCRIFX PROJECT DESCRIPTION - DRAFT (microarrays)

Project Number: CCRIFX-

Project Title:

Requestor:

Requestor Lab PI:

Address:

Additional Investigators to include:

Bioinformatics contact:

Completed Request(s):

Current Active Request(s):

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### Rationale/Significance:

#### Scientific questions or goals (choose all that apply)

- **Transcriptional profiling:** aimed to estimate absolute genes expression values in a cell type/tissue
- **Identification of gene expression signatures:** aimed to identify expression patterns uniquely characteristic of a medical or other condition such as tumor stage, subtype, aggressiveness, metastatic potential, etc.
- **Class comparison:** aimed to determine whether gene expression profiles differ among samples from different classes and to identify which genes are differentially expressed between classes.
- **Testing for interaction between classes:** aimed to identify synergistic effects between two factors (e.g. two drugs).
- **Class prediction:** aimed to develop a statistical model that can predict which class a new specimen belongs to based on its expression profile.
- **Class discovery:** aimed to classify samples and/or to identify novel subtypes of specimens within a population. E.g. which gene expression signatures are associated with prolonged survival?
- **Enrichment analysis:** aimed to identify differentially regulated pathways
- **Gene network reconstruction:** aimed to do reverse engineering of gene regulatory networks

### Approach

- *E.g. Microarray gene expression profiling of murine fibroblasts (KO and WT) treated with ...*

### Impact

- *If successful, this study will identify the role of XXX in immune response...*

### Other priority considerations

- Event(s) in the near future that makes this request time-sensitive:
  - Site visits
  - Manuscripts/Proposals
  - Posters/Presentations
- The availability of data to be analyzed:

- Not available yet
  - Available now
  - Public sources
- Role of the project:
  - Publishable experiment
  - Exploratory
  - Confirmatory

## Known risks and limitations

Gene expression microarray profiling projects can have dramatically different goals. To maximize the chance of meeting the goals, it is critical to use appropriate experimental design.

- The following factors can help reduce bias and variability in microarray experiments:
  - Ideally sample collection and NA extraction should be performed the same day by the same individual using the same protocol and reagents.
  - If that is not possible, samples should be frozen and processed together at a later date at the same microarray facility. For large studies, randomization and blocking should be used when applicable.
- At least three replicates per condition are recommended to allow for statistical analysis. More is better, because it will improve the sensitivity of the analysis.
- More replicates are recommended for experiments that involve
  - High biological variability (e.g. human or mouse samples vs. cell lines)
  - Non-target tissue contamination (e.g. mouse embryonic tissues)
  - Subtle treatment effect (e.g. weak transcriptional signal)
  - Multiple treatments
  - The goals include understanding the mechanism of action or network analysis
- Note: For more guidance, see the checklist for Nature journals:
  - [www.nature.com/authors/policies/checklist.pdf](http://www.nature.com/authors/policies/checklist.pdf)

## Original description

*[Text describing the analysis goes here]*

## Experimental design and metadata

- **Samples**
  - Species and strains:
    - Human (race if known)
    - Mouse (e.g. C57BL/6, FVB/NJ, 129P2/OlaHsd)
    - Other (please specify)
  - Type: tissue, cell lines, cells, etc.
  - Extracted material: non/polyadenylated long RNA, small RNA, miRNA, etc.
  - Total number of
    - Samples -
    - Groups -

- **Replicates (biological and technical) -**

- Diversity of the population, intra-individual variability of the sample, clonal heterogeneity, similarity to the reference, non-target tissue contamination, etc. – if known

- **Protocols**

- GEO submission fields (Excel file)
- Treatment and enrichment protocols used in sample prep:
- Microarray platforms used:
  - Affymetrix
  - cDNA or spotted arrays
  - Other one-color arrays: Agilent, Applied Biosystems, Eppendorf, GE Healthcare, Illumina, Nimblegen
  - Other two-color arrays: Agilent, NCI\_Operon
  - Pathway-Specific PCR arrays

- **Data**

- Location and format: cel, chp and grd files
- **SAIC-F CSAS#**
- Preferred genome references: version/source (e.g. hg19, mm9, mm10)
- For public data mining requests:
  - Diseases, cancer subtypes, databases: ENCODE, GEO, etc.
  - Types of data: RNA-Seq, ChIP-Seq, microarray, etc.

## **Analysis Details**

- **Analysis desired from CCRIFX**

- Preferred analytical applications – standard or custom
- Major steps in the workflow or procedure that will be followed
  - Normalization and expression value estimation
  - QC: PCA and Correlation plots, R coefficient across replicates
  - QC tools: Affymetrix Power Tools; R/Bioconductor – Simpleaffy or arrayQualityMetrics (platform agnostic)
- Differentially expressed genes (DEG) analysis
  - Software tools
    - BRB Arraytools
    - GeneSpring
    - Partek
    - LIMMA package
    - Affymetrix GCOS
  - Comparisons and controls
    - DE analysis
    - Testing for interaction between factors
- Genome features of interest: genes, transcripts, exons

- **Results expected from CCRIFX**

- Files: gene lists, pathway lists, etc.
- Figures: heatmaps, venn diagrams
- List of methods used with brief descriptions

- Education and training
- Submission to public repositories: GEO, GenBank

**Deferred tasks**

- Related analyses on the same data that are too big to fit into the current request
- Related analyses on that require additional data (e.g. data integration)
- Requests for continuous support

**Publications**

- References specific approaches, analyses or protocols that the investigator wants to replicate
- Publications that provide scientific context for the analysis